

4. Wang G, van Dam AP, Dankert J. Phenotypic and genetic characterization of a novel *Borrelia burgdorferi* sensu lato isolate from a patient with Lyme borreliosis. *J Clin Microbiol.* 1999;37:3025–8.
5. Fingerle V, Michel H, Schulte-Spechtel U, Göttner G, Hizo-Teufel C, Hofmann H, et al. A14S—a new *Borrelia burgdorferi* s.l. genospecies as relevant cause of human disease [abstract]. *Int J Med Microbiol.* 2004;294(Suppl 1):207.
6. Rauter C, Oehme R, Dietrich I, Engele M, Hartung T. Distribution of clinically relevant *Borrelia* genospecies in ticks assessed by a novel, single-run real-time PCR. *J Clin Microbiol.* 2002;40:36–43.
7. Michel HB, Wilske B, Hettche G, Göttner G, Heimerl C, Reischl U, et al. An ospA-polymerase chain reaction/restriction fragment length polymorphism-based method for sensitive detection and reliable differentiation of all European *Borrelia burgdorferi* sensu lato species and OspA types. *Med Microbiol Immunol.* 2003;193:219–26.
8. Derdákóvá M, Beati L, Pet'ko B, Stanko M, Fish D. Genetic variability within *Borrelia burgdorferi* sensu lato genospecies established by PCR-single-strand conformation polymorphism analysis of the rrfA-rrlB intergenic spacer in *Ixodes ricinus* ticks from the Czech Republic. *Appl Environ Microbiol.* 2003;69:509–16.
9. Richter D, Schlee DB, Allgöwer R, Matuschka FR. Relationships of a novel Lyme disease spirochete, *Borrelia spielmani* sp. nov., with its hosts in central Europe. *Appl Environ Microbiol.* 2004;70:6414–9.
10. Demaerschalck I, Benmessaoud A, Dekesel M, Hoyois B, Lobet Y, Hoet P, et al. Simultaneous presence of different *Borrelia burgdorferi* genospecies in biological fluids of Lyme-disease patients. *J Clin Microbiol.* 1995;33:602–8.

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Profiling *Mycobacterium ulcerans* with *hsp65*

To the Editor: *Mycobacterium ulcerans* is an emerging human pathogen responsible for Buruli ulcer, a necrotizing skin disease most commonly found in West Africa, but outbreaks have also been reported in the Americas, Australia, and Asia (1). Environmental sources of infection and mode of transmission are not completely known. *M. ulcerans* grows slowly at 32°C, requiring 6–8 weeks for colonies to be visible in primary culture. Differentiation from *M. marinum*, which also causes skin infections, is important, since *M. marinum* can usually be treated with antimicrobial agents, whereas *M. ulcerans* most often does not respond favorably to drug therapy, and treatment is usually by surgical excision (2). *M. shinshuense*, initially isolated from a child in Japan, is phenotypically and genetically related but biochemically distinct from *M. ulcerans* (3).

In the last decade, several DNA-based techniques for mycobacterial identification have been developed. Rapid molecular detection and differentiation of organisms that cause skin infections directly from tissue or exudates could be of great value for early treatment. Some techniques, especially those that include nucleic acid amplification, could be used directly on clinical samples. The accepted standard for molecular identification of mycobacteria is sequencing analysis of 2 hypervariable regions identified in 16S rRNA gene. *M. marinum* and *M. ulcerans* share identical 5′-16S rDNA and 16S-23S rRNA gene spacer sequences (4). Polymerase chain reaction (PCR)-dependent methods are based on the 16S rRNA gene (5), the *hsp65* gene (6) or the insertion sequence IS2404 (7).

Recently, a novel category of variable number tandem repeats that could distinguish *M. marinum* and *M. ulcerans* genotypes has been described (8).

Polymorphisms in the 3′-16S rDNA region discriminate *M. ulcerans* from *M. marinum* and *M. shinshuense* (5). These polymorphisms also allow the separation of *M. ulcerans* into 3 subgroups according to geographic origin and variable phenotypic differences. IS2404 discriminates *M. ulcerans* from *M. marinum* (7). It has been used in restriction fragment length polymorphism analysis applied to a comparable number of *M. ulcerans* and *M. marinum* strains, confirming that this sequence is present in high copy numbers in *M. ulcerans* but absent in *M. marinum*. Nevertheless, an unusual mycobacterium was recently isolated that is closely related to *M. marinum* by phenotypic tests, lipid pattern, and partial 16S rDNA sequencing but presents low copy numbers of this element (9).

PCR-restriction enzyme analysis (PRA) of a 441-bp fragment of the *hsp65* gene is a rapid, easy, and inexpensive method for identifying mycobacteria (10). Devallois et al. (6) described the PRA-*hsp65* pattern of 1 *M. ulcerans* strain ATCC 33728 that originated in Japan. This isolate was considered a new species that resembled *M. ulcerans* and was named *M. shinshuense* (3).

We report here the usefulness of PRA-*hsp65* to differentiate *M. ulcerans* strains from different geographic areas. Since Buruli ulcer cases have been reported on 5 continents, we studied 33 *M. ulcerans* strains that originated from Africa (Benin, Zaire, Ghana, Congo, Angola, Côte d'Ivoire, Togo), Asia (China, Malaysia), Australia (Papua New Guinea, Australia), the Caribbean (Mexico, Surinam, French Guiana), 1 *M. shinshuense* from Japan, 1 *M. marinum* isolate and 1 IS2404-positive *M. marinum* isolate from France (9). All strains were identified at the Institute

of Tropical Medicine, the World Health Organization Collaborating Centre for the Diagnosis and Surveillance of *Mycobacterium ulcerans* Infection by IS2404 PCR and biochemical tests (online Table, available from <http://www.cdc.gov/ncidod/EID/vol11no11/05-0234.htm#table>).

DNA extracted from cultures by 3 freeze-boiling cycles was used for amplification, according to the protocol described by Leao et al. (10). Gel images were analyzed by using GelCompar II v. 2.5 (AppliedMaths, Sint-Martens-Latem, Belgium). Two distinct *M. ulcerans* PRA-*hsp65* patterns were identified. Of 36 strains, 34 had a PRA-*hsp65* pattern indistinguishable from that of *M. marinum* [*Bst*EII and *Hae*III (bp) of 235/210/0 and 145/105/80] at the Swiss PRA site (<http://app.chuv.ch/prasite/index.html>). Two strains, 1 each from Japan and China, showed a different pattern [*Bst*EII and *Hae*III (bp) of 235/210/0 and 190/105/80], that described by Devallois et al. (6).

We have shown that PRA-*hsp65* analysis performed on several *M. ulcerans* strains from different geographic areas produced different patterns. In fact, the unique PRA-*hsp65* profile of the *M. ulcerans* strain previously published (6) was the most rarely found pattern among the profiles found in this study. This work helps to clarify the PRA-*hsp65* patterns of *M. ulcerans* found in different countries. Because the epidemiology of Buruli ulcer is poorly understood, new molecular tools are still needed to differentiate *M. ulcerans* from different geographic settings, mainly in Africa, where the disease is more prevalent. The PRA-*hsp65* method represents a rapid, easy, and inexpensive technique to differentiate *M. shinshuense* from *M. ulcerans* and *M. marinum*.

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References

- Asiedu K, Scherpier R, Raviglione M, editors. Buruli ulcer. *Mycobacterium ulcerans* infection. Geneva: The World Health Organization; 2000.
- Buntine J, Crofts K, editors. Buruli ulcer. Management of *Mycobacterium ulcerans* disease. Geneva: The World Health Organization; 2001.
- Tsukamura M, Kaneda K, Imaeda T, Mikoshiha H. [A taxonomic study on a mycobacterium which caused a skin ulcer in a Japanese girl and resembled *Mycobacterium ulcerans*]. *Kekkaku*. 1989;64:691–7.
- Roth A, Fischer M, Hamid ME, Michalke S, Ludwig W, Mauch H. Differentiation of phylogenetically related slowly growing mycobacteria based on 16S-23S rRNA gene internal transcribed spacer sequences. *J Clin Microbiol*. 1998;36:139–47.
- Portaels F, Fonteyne PA, de Beenhouwer H, de Rijk P, Guedenon A, Hayman J, et al. Variability in 3' end of 16S rRNA sequence of *Mycobacterium ulcerans* is related to geographic origin of isolates. *J Clin Microbiol*. 1996;34:962–5.
- Devallois A, Goh KS, Rastogi N. Rapid identification of mycobacteria to species level by PCR-restriction fragment length polymorphism analysis of the *hsp65* gene and proposition of an algorithm to differentiate 34 mycobacterial species. *J Clin Microbiol*. 1997;35:2969–73.
- Stinear T, Ross BC, Davies JK, Marino L, Robins-Browne RM, Oppedisano F, et al. Identification and characterization of IS2404 and IS2606: two distinct repeated sequences for detection of *Mycobacterium ulcerans* by PCR. *J Clin Microbiol*. 1999;37:1018–23.
- Stragier P, Ablordey A, Meyers WM, Portaels F. Genotyping *Mycobacterium ulcerans* and *Mycobacterium marinum* using mycobacterial interspersed repetitive units. *J Bacteriol*. 2005;187:1639–47.
- Chemlal K, Huys G, Laval F, Vincent V, Savage C, Gutierrez C, et al. Characterization of an unusual *Mycobacterium*: a possible missing link between *Mycobacterium marinum* and *Mycobacterium ulcerans*. *J Clin Microbiol*. 2002;40:2370–80.
- Leao SC, Bernardelli A, Cataldi A, Zumarraga M, Robledo J, Realpe T, et al. Multicenter evaluation of mycobacteria identification by PCR restriction enzyme analysis in laboratories from Latin America and the Caribbean. *J Microbiol Methods*. 2005;61:193–9.

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Spelling of Emerging Pathogens

To the Editor: Language is about comprehension; provided the parties in a discussion can understand each other, variations in pronunciation of individual words may be tolerated or disregarded. In modern English, numerous examples of variant pronunciations exist that cause no problems of comprehension (e.g., either, tomato, laboratory, fertile). These arise from several causes; regional practice is likely the most important factor, but the speaker's education and social background, personal preferences, and even etymologic theories also play a part. It would be futile and, some would feel, undesirable to attempt to impose uniformity by prescribing approved pronunciations if communication is not endangered. Moreover, both language and pronunciation are subject to constant change.

The same is not true regarding the spelling of organisms' names.